

New Quassinoids from the Roots of *Eurycoma longifolia*Hiroshi MORITA, Etsuko KISHI, Koichi TAKEYA, Hideji ITOKAWA,* and Osamu TANAKA[†]

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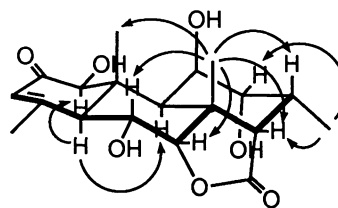
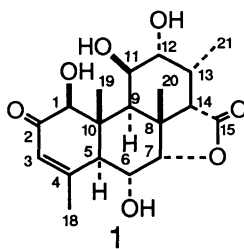
A new C₁₉-skeleton quassinoid, named longilactone and three new quassinoids, 13,21-dihydroeurycomanone, 13 β ,21-dihydroxyeurycomanone and 14,15 β -dihydroxyklaineane were isolated from the roots of *Eurycoma longifolia* and their structures were established by the spectral evidences.

From *Eurycoma longifolia* Jack (Simaroubaceae) that is one of famous folk medicines in the Southeast Asia, C₂₀-quassinoids; eurycomanone,¹⁾ eurycomanol,¹⁾ eurycomanol-2-*O*- β -D-glycopyranoside,²⁾ C₁₉-; eurycomalactone whose structure was revised to form a lactonic linkage at not C-7 but C-12,³⁾ C₁₈-; laurycolactones A and B³⁾ were so far isolated and some of them exhibited *in vitro* antimalarial activities.⁴⁾ Further investigation of the *n*-butanol extract resulted in isolation of new cytotoxic quassinoids, named longilactone (**1**) which is a new C₁₉-type quassinoid to form a lactonic linkage at C-7, 13,21-dihydroeurycomanone (**2**), 13 β ,21-dihydroxyeurycomanone (**3**) and 14,15 β -dihydroxyklaineane (**4**) along with known eurycomanone (**5**) and eurycomanol (**6**). In this communication, the structure elucidation of them and cytotoxic activities are described.

The roots of *E. longifolia* collected in Indonesia were extracted with 50% methanol, and chromatographic purification of *n*-butanol soluble fraction furnished compounds **1-6**.

Longilactone (**1**)⁵⁾ was suggested to be C₁₉ quassinoid by ¹³C-NMR spectrum and the molecular formula, C₁₉H₂₆O₇ was confirmed by high resolution MS spectrum (366.1675). The IR and UV spectra showed the presence of an α,β -unsaturated ketone (1656 cm⁻¹; 242 nm, ϵ =10500) and a butanolide (1769 cm⁻¹) like eurycomalactone (**7**).³⁾ Detailed decoupling experiments based on ¹H-¹H and ¹H-¹³C COSY spectra in 500MHz NMR revealed us the complete assignments and coupling network as listed in Tables 1 and 2. Compared with the NMR data of **7**, the signal of H-12 (δ 3.60) and also the signal of the corresponding carbon (δ 76.2) showed high field shift. Furthermore, the ¹H-¹H coupling constants around C-ring were different between **1** and **7**, for example, the coupling constant (1.0Hz) between H-12 and H-14 and negligible coupling constant (ca.0Hz) between H-12 and H-13 characteristic of **7** were not observed in **1**. On the other hand, the appearance of oxygen-substituted functions at C-6 and C-7 was secured by the coupling sequence from H-5. Then, judging by the chemical shift (δ 66.0 and 88.1) of these carbons and the presence of ¹H-¹³C long rang coupling between H-7 and C-15, it turned out to substitute a hydroxyl group at C-6 and a lactonic linkage at C-7. The above all data are consistent with the proposed structure with a lactonic linkage between C-7 and C-15.

The stereostructure was corroborated by difference NOE experiments in ^1H -NMR. Irradiation of H-20 caused increase in the integrated intensity of H-19, H-6, H-7, H-13, and H-14, respectively. Then, NOE among H-1, H-5, and H-9 was observed as shown in Figure. Further, from the coupling constants, the structure was also explained satisfactorily.

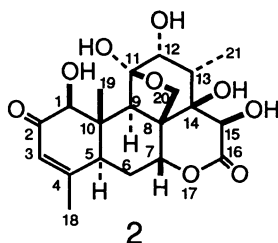


NOE enhancements

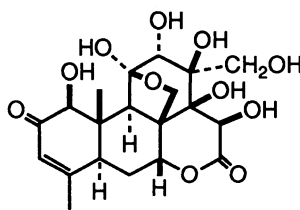
Compounds **2**⁶⁾ and **3**⁷⁾ were obtained as colorless needles, respectively and were presumed to be C_{20} quassinoids by ^{13}C -NMR spectrum. The molecular formulae of **2** and **3** were established to be $\text{C}_{20}\text{H}_{26}\text{O}_9$ (410.1582) and $\text{C}_{20}\text{H}_{26}\text{O}_{11}$ by high resolution MS and FAB MS spectra and the structures of them were considered to be closely related structures to **5** by spectral data. The IR and UV spectra indicated the presence of an α,β -unsaturated ketone (**2**; 1680 cm^{-1} and 235 nm , **3**; 1672 cm^{-1} and 240 nm) and a lactone (**2**; 1740 cm^{-1} , **3**; 1728 cm^{-1}). In ^1H -NMR spectrum, the signals of a doublet methyl ($\delta 1.85$) and a methine ($\delta 2.84$) coupling with the methyl were newly appeared in **2** in place of those ascribable to exomethylene in **5**. The carbon peaks ($\delta 44.2$ and 14.0) corresponding to this unit were also present. In the case of **3**, the presence of 1,2-glycol at C-13 and C-21 was implied by the carbon signals at $\delta 77.9$ and 66.8 . The data described above led us to elucidate **2** and **3** as shown in Figure. The stereostructure was proved by difference NOE experiments, the integrated intensity between H-13 and H-20 in **2** and among H-21, H-12, and H-15 in **3** were increased.

Compound **4**⁸⁾ was isolated as colorless needles, mp $134\text{--}135^\circ\text{C}$, $\text{C}_{20}\text{H}_{28}\text{O}_8$ (378.1638, $\text{M}^+ - \text{H}_2\text{O}$). The NMR data of **4** suggested that the acetal linkage between C-20 and C-11 in **2** is not present. Then, configurations of hydroxyl groups at C-11, 12, and 15 were determined by the ^1H -coupling constants similar to those of **1** and NOE that observed between H-20 and H-13, H-21 and H-12, and then H-9 and H-15. Absolute structures of **2**, **3** and **4** were determined to be same as **5** because of the appearance of analogous cotton curve⁶⁻⁸⁾ to **5**.

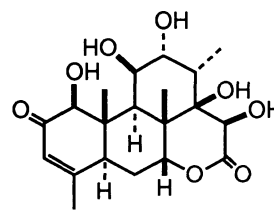
Compounds **5**⁹⁾ and **6**¹⁰⁾ were identical with eurycomanone and eurycomanol, respectively by comparison with various spectral data.



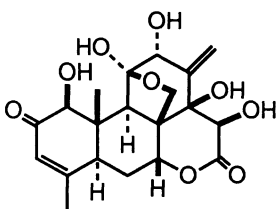
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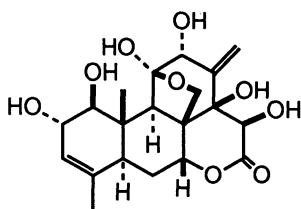
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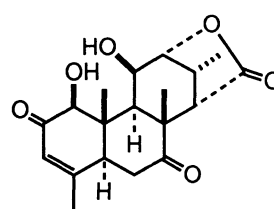
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5



6



7

Table 1. ¹H-NMR spectral data for 1-7 (500 MHz)

	1a)	2b)	3b)	4a)	5b)	6b)	7c)	
H-1	3.99,s	4.39,s	4.45,s	4.08,s	4.52,s	4.04,d	4.05,brs	Coupling constants in Hz
H-2	---	---	---	---	---	4.63,m	---	
H-3	6.05,brs	6.13,brs	6.13,brs	6.04,brs	6.16,brs	5.80,brs	6.11,brs	1: J _{5,6} =11.0; J _{6,7} =3.9; J _{9,11} =2.0; J _{11,12} =3.0;
H-5	2.75,brd	3.17,brd	3.19,brd	2.92,brd	3.26,brd	2.82,brd	2.80,m	J _{12,13} =3.0; J _{13,14} =3.0; J _{13,21} =7.0
H-6e	---	2.27,dt	2.27,dt	2.22,dt	2.34,dt	2.19,dt	2.65,dd	2: J _{5,6e} =2.5; J _{5,6a} =14.0; J _{6e,6a} =14.0; J _{6e,7} =
H-6a	4.21,dd	2.03,td	2.07,td	2.14,td	2.03,td	1.93,td	2.75,dd	2.5; J _{6a,7} =2.5; J _{12,13} =4.1; J _{13,21} =7.3;
H-7	4.12,d	5.21,t	5.12,t	4.52,t	5.26,t	5.16,t	---	J _{20,20} =9.0
H-9	2.05,d	3.51,s	3.59,s	2.16,d	3.82,s	3.53,s	1.86,d	3: J _{5,6e} =2.5; J _{5,6a} =14.0; J _{6e,6a} =14.0; J _{6e,7} =
H-11	4.95,dd	---	---	5.02,t	---	---	4.78,brdd	2.5; J _{6a,7} =2.5; J _{12,OH} =4.6; J _{21,21} =11.6;
H-12	3.60,t	4.14,d	4.58,d	3.68,t	4.80,s	4.78,s	4.38,brd	J _{20,20} =8.8
H-13	2.35,m	2.84,m	---	2.36,qd	---	---	2.80,m	4: J _{5,6e} =3.0; J _{5,6a} =14.7; J _{6e,6a} =14.7; J _{6e,7} =
H-14	2.34,d	---	---	---	---	---	2.88,d	3.0; J _{6a,7} =3.0; J _{9,11} =3.0; J _{11,12} =3.0;
H-15	---	5.62,s	5.57,s	5.41,s	5.67,s	5.49,s	---	J _{12,13} =3.0; J _{13,21} =7.3
H-18	2.28,brs	1.77,brs	1.78,brs	1.99,brs	1.80,brs	1.64,brs	1.95,brs	5: J _{5,6e} =2.7; J _{5,6a} =14.0; J _{6e,6a} =14.0; J _{6e,7} =
H-19	1.09,s	1.62,s	1.65,s	1.21,s	1.62,s	1.74,s	1.26,s	2.7; J _{6a,7} =2.7; J _{21,21} =1.4; J _{20,20} =8.6
H-20	1.57,s	4.04,d	3.97,d	1.50,s	4.01,d	4.05,d	1.56,s	6: J _{1,2} =7.8; J _{5,6e} =2.5; J _{5,6a} =14.0; J _{6e,6a} =14.0
		4.64,d	5.20,d		4.54,d	4.55,d		J _{6e,7} =2.5; J _{6a,7} =2.5; J _{21,21} =1.5; J _{20,20} =8.5
H-21	1.38,d	1.85,d	4.61,d	1.23,d	5.66,d	5.63,d	1.16,d	7: J _{5,6e} =4.0; J _{5,6a} =13.5; J _{6e,6a} =15.5; J _{9,11} =
			5.01,d		6.12,d	6.09,d		3.5; J _{11,12} =5.0; J _{12,13} =ca. 0.0; J _{12,14} =1.0;
OH-12			7.86,d					J _{13,21} =7.0

a) In CD₃OD b) in pyridine-d₅ c) in CDCl₃, cited from Ref. 3.Table 2. ¹³C-NMR spectral data for 1-7 (100 MHz)

	C-1	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10	C-11	C-12
1a)	83.6	201.2	127.4	167.7	51.6	66.0	88.1	44.2	42.9	50.9	73.7	76.2
2b)	84.6	197.5	126.1	162.7	42.2	25.8	71.9	52.7	47.0	45.7	110.3	79.7
3b)	84.7	197.5	126.1	162.4	42.3	25.7	70.6	53.5	47.4	45.5	110.0	79.6
4b)	84.8	199.4	124.8	165.0	43.4	26.3	71.2	49.2	44.7	44.1	74.3	80.9
5b)	84.6	197.4	126.1	162.5	42.2	25.7	71.8	52.6	47.7	45.9	109.6	81.0
6b)	83.8	72.7	126.9	135.0	41.6	25.6	71.9	52.7	47.9	42.3	109.7	81.1
7c)	81.3	197.4	124.5	162.2	49.5	36.3	205.5	47.0	53.0	51.2	69.9	83.2

	C-13	C-14	C-15	C-16	C-18	C-19	C-20	C-21	
1	28.2	57.1	178.9	---	21.3	12.7	25.3	14.8	a) in CD ₃ OD
2	42.2	76.6	75.1	174.7	22.4	10.6	67.3	14.0	
3	77.9	78.3	74.9	173.5	22.4	11.0	67.6	66.8	b) in Pyridine-d ₅
4	35.7	77.5	78.0	175.9	18.2	12.0	22.4	14.2	
5	148.0	79.4	75.9	173.8	22.4	10.4	67.7	119.3	c) in CDCl ₃ , cited from Ref. 3
6	148.5	79.4	76.5	173.8	21.2	10.9	67.8	119.3	
7	32.4	49.2	176.2	---	22.0	12.2	23.7	16.7	

Compounds **1-6** exhibited cytotoxic activities against KB and P-388 cells.¹¹⁾ It seems that saturation at C-13 and C-21 such as **1**, **2**, and **4** may be structural requirement for potent activity.

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References

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- 4) K. L. Chan, M. J. O'Neill, J. D. Phillipson, and D. C. Warhurst, *Planta Medica*, 105 (1986).
- 5) Longilactone (**1**): mp 130-132 °C, $[\alpha]_D^{+92.6^\circ}$ (c 0.19, MeOH). CD (EtOH): $[\theta]_{240} +22300$.
- 6) 13,21-dihydroeurycomanone (**2**): mp 219-221 °C, $[\alpha]_D^{+46.9^\circ}$ (c 0.11, MeOH). CD (EtOH): $[\theta]_{240} -9400$, $[\theta]_{322} +1970$.
- 7) 13 β ,21-dihydroxyeurycomanone (**3**): mp 229-230 °C, $[\alpha]_D^{+17.5^\circ}$ (c 0.16, pyridine). CD (EtOH): $[\theta]_{245} -12900$, $[\theta]_{320} +1840$.
- 8) 14,15 β -dihydroxyklaineanone (**4**): mp 134-135 °C, $[\alpha]_D^{+53.8^\circ}$ (c 0.29, MeOH). UV (EtOH): 240 nm (ϵ 8510). CD (EtOH): $[\theta]_{247} -9900$, $[\theta]_{315} +2150$.
- 9) Eurycomanone (**5**): mp 242-243 °C, $[\alpha]_D^{+34.2^\circ}$ (c 0.32, pyridine).
- 10) Eurycomanol (**6**): mp 258-260 °C, $[\alpha]_D^{+85.6^\circ}$ (c 0.31, pyridine).
- 11) Cytotoxic activities against KB and P-388 cells (IC₅₀ μ g/ml)

	Compound 1	2	3	4	5	6
KB Cells	3.4	0.33	20	0.38	0.40	3.6
P-388 Cells	1.3	1.2	10	0.29	10	58

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